

RESEARCH ARTICLE

# Synthesis, spectroscopic, and antibacterial activity of tetraazamacrocyclic complexes of trivalent chromium, manganese, and iron

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## Abstract

A new series of macrocyclic complexes of type  $[M(\text{TML})\text{X}]_2$ , where  $M = \text{Cr(III)}$ ,  $\text{Mn(III)}$ , or  $\text{Fe(III)}$ , TML is tetradentate macrocyclic ligand, and  $\text{X} = \text{Cl}^-$ ,  $\text{NO}_3^-$ ,  $\text{CH}_3\text{COO}^-$  for  $\text{Cr(III)}$ ,  $\text{Fe(III)}$  and  $\text{X} = \text{CH}_3\text{COO}^-$  for  $\text{Mn(III)}$ , has been synthesized by condensation of benzil and succinyldihydrazide in the presence of metal salt. The complexes have been so formulated due to the 1:2 electrolytic nature of these complexes as shown by conductivity measurements. The complexes have been characterized with the help of various physicochemical techniques such as elemental analysis, molar conductance, electronic and infrared spectral studies, and magnetic susceptibility. On the basis of these studies, a five-coordinate distorted square pyramidal geometry, in which two nitrogens and two carbonyl oxygen atoms are suitably placed for coordination toward the metal ion, has been proposed for all the complexes. The complexes have been tested for their *in vitro* antibacterial activity. Some of the complexes show remarkable antibacterial activities against some selected bacterial strains. The minimum inhibitory concentrations shown by these complexes have been compared with those shown by some standard antibiotics such as linezolid and cefaclor.

**Keywords:** Macrocyclic complexes; template; infrared; ligand; antibacterial

**Abbreviations:** MIC, minimum inhibitory concentration; MTCC, microbial type culture collection; MHA, Muller–Hinton agar; CFU, colony forming unit; BM, Bohr magneton; DMF, N,N-dimethylformamide; DMSO, dimethyl sulfoxide; BHI, brain heart infusion

## Introduction

Transition metals play an important role in the development of new molecular materials which show magnetic properties and find applications in various fields. During the past few decades, the synthesis of macrocyclic complexes has been a fascinating area of research growing at a very fast pace, owing to their resemblance to naturally occurring macrocycles and analytical, industrial, and medical applications<sup>1–5</sup>. Macrocyclic nickel complexes find use in DNA recognition and oxidation<sup>6</sup> while macrocyclic copper complexes find use in DNA binding and cleavage<sup>7</sup>. Macrocyclic metal complexes of lanthanides, e.g.  $\text{Gd}^{3+}$  are used as magnetic resonance imaging (MRI) contrast agents<sup>8,9</sup>. Macrocyclic metal chelating agents (DOTA) are

useful for detecting tumor lesions<sup>10</sup>. The chemistry of macrocyclic complexes is also important due to their use as dyes and pigments<sup>11</sup> as well as nuclear magnetic resonance (NMR) shift reagents<sup>12</sup>. Some macrocyclic complexes have been found to exhibit potential antibacterial activities<sup>13</sup>. Prompted by these, in the present article a new series of macrocyclic complexes of  $\text{Cr(III)}$ ,  $\text{Fe(III)}$ , and  $\text{Mn(III)}$  obtained by template condensation reaction of succinyldihydrazide and benzil is reported. The complexes have been characterized with the help of infrared (IR) and magnetic susceptibilities, elemental analysis, and molar conductance. These complexes have also been tested for their *in vitro* antibacterial activities. Some complexes show remarkable antibacterial activity.

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## Experimental

### Chemistry

#### Materials

Diethylsuccinate, hydrazine hydrate, and benzil were obtained from SRL Pvt. Ltd., Mumbai. All other chemicals used were of AnalR grade.

#### Analytical and physical measurements

Microanalyses of C, H, and N were carried out at the Sophisticated Analytical Instrument Facility, CDRI, Lucknow. The metal contents were determined by standard ethylenediaminetetraacetic acid (EDTA) methods. Electronic spectra (DMF) were recorded on a Cary 14 spectrophotometer. The magnetic susceptibility measurements were carried at IIT, Roorkee. The IR spectra were recorded on an infrared spectrophotometer in the range  $4000\text{--}200\text{ cm}^{-1}$ . The conductivity was measured using a digital conductivity meter (HPG System, G-3001).

#### Isolation of complexes

All the reported complexes were prepared using the template method. To a stirring methanolic solution ( $\sim 50\text{ cm}^3$ ) of succinyldihydrazide (10 mmol) was added trivalent chromium, manganese, and iron salt (10 mmol) dissolved in a minimum quantity of methanol ( $20\text{ cm}^3$ ). The resulting solution was refluxed for 0.5 h. After that, benzil (10 mmol) dissolved in  $\sim 20\text{ mL}$  of methanol was added to the refluxing mixture which was refluxed again for 6–8 h. On overnight cooling, a dark colored precipitate formed, which was filtered, washed with methanol, acetone, and diethyl ether, and dried *in vacuo* (yield 45%). The complexes were found to be soluble in DMF and DMSO, but were insoluble in common organic solvents and water. They were found to be thermally stable up to  $\sim 240^\circ\text{C}$  and then they decomposed.

The template syntheses of the complexes may be represented by the scheme shown in Figure 1, where  $M = \text{Cr(III)}$ ,  $\text{Mn(III)}$ ,  $\text{Fe(III)}$ , and  $X = \text{Cl}^-$ ,  $\text{NO}_3^-$ ,  $\text{CH}_3\text{COO}^-$  for  $\text{Cr(III)}$  and  $\text{Fe(III)}$  and  $\text{CH}_3\text{COO}^-$  for  $\text{Mn(III)}$ .

### Pharmacology

#### In vitro antibacterial activity

Some of the synthesized macrocyclic complexes were tested for *in vitro* antibacterial activity against some bacterial strains using spot-on-lawn on Muller-Hinton agar by a reported method<sup>14</sup>.

**Test pathogens** Four test pathogenic bacterial strains, viz. *Bacillus cereus* (MTCC 1272), *Salmonella typhi* (MTCC 733), *Escherichia coli* (MTCC 739), and *Staphylococcus aureus* (MTCC 1144), were considered for determination of the MIC (minimum inhibitory concentration) of selected complexes.

**Culture conditions** The test pathogens were subcultured aerobically using brain heart infusion agar (HiMedia, Mumbai, India) at  $37^\circ\text{C}/24\text{ h}$ . Working cultures were stored at  $4^\circ\text{C}$  in brain heart infusion (BHI) broth (HiMedia), while stock cultures were maintained at  $-70^\circ\text{C}$  in BHI broth containing 15% (v/v) glycerol (Qualigens, Mumbai, India). Organism was grown overnight in 10 mL BHI broth, which was then centrifuged at  $5000g$  for 10 min, and the pellet was suspended in 10 mL of phosphate buffered saline (PBS, pH 7.2). The optical density at 545 nm (OD-545) was adjusted to obtain  $10^8$  CFU/mL, followed by plating serial dilution onto plate count agar (HiMedia).

**Determination of minimum inhibitory concentration** The minimum inhibitory concentration (MIC) is the lowest concentration of the antimicrobial agent that prevents the development of viable growth after overnight incubation. The antimicrobial activity of the compounds was evaluated using the spot-on-lawn method on Muller-Hinton agar (MHA; HiMedia). Soft agar was prepared by adding 0.75% agar in Muller-Hinton broth (HiMedia). The soft agar was inoculated with 1% of  $10^8$  CFU/mL of the test pathogen, and 10 mL was overlaid on MHA. From a  $\times 1000$  solution of compound (1 mg/ml of DMSO),  $\times 1$ ,  $\times 2$ ,  $\times 4$ ,  $\times 8$ ,  $\times 16$ ,  $\times 32$ ,  $\times 64$ , and  $\times 128$  solutions were prepared. Dilutions of standard antibiotics (linezolid and cefaclor) were also prepared in the same manner. Five microliters of the appropriate dilution was spotted on the soft agar and incubated at  $37^\circ\text{C}$  for 24 h. Zones of inhibition of the compounds were considered after subtraction of the inhibition zones of DMSO. A negative control (with no compound) was also observed.

## Results and discussion

### Chemistry

The analytical data show the formula of macrocyclic complexes as:  $[\text{M}(\text{C}_{18}\text{H}_{16}\text{N}_4\text{O}_2)\text{X}]_2$ , where  $M = \text{Cr(III)}$ ,  $\text{Mn(III)}$ ,  $\text{Fe(III)}$  and  $X = \text{Cl}^-$ ,  $\text{NO}_3^-$ ,  $\text{CH}_3\text{COO}^-$  for  $\text{Cr(III)}$  and  $\text{Fe(III)}$  and  $X = \text{CH}_3\text{COO}^-$  for  $\text{Mn(III)}$ . The test for anions was positive before and after decomposing the complexes with

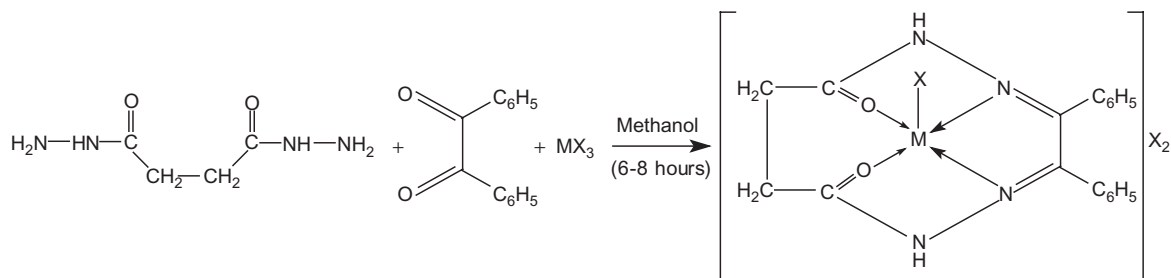


Figure 1. Synthesis of the complexes.

**Table 1.** Analytical data for trivalent chromium, manganese, and iron complexes derived from succinyldihydrazide and benzil.

Series no.	Complex	Found (calculated) (%)				Color	Mol. wt.
		M	C	H	N		
1	[Cr(C <sub>18</sub> H <sub>16</sub> N <sub>4</sub> O <sub>2</sub> )Cl]Cl <sub>2</sub>	10.88 (10.86)	45.21 (45.14)	3.29 (3.34)	11.79 (11.70)	Light green	478.5
2	[Cr(C <sub>18</sub> H <sub>16</sub> N <sub>4</sub> O <sub>2</sub> )(NO <sub>3</sub> )](NO <sub>3</sub> ) <sub>2</sub>	9.23 (9.31)	38.67 (38.70)	2.95 (2.86)	17.62 (17.56)	Light yellow	558
3	[Cr(C <sub>18</sub> H <sub>16</sub> N <sub>4</sub> O <sub>2</sub> )(OAc)](OAc) <sub>2</sub>	9.49 (9.47)	52.53 (52.45)	4.48 (4.55)	10.23 (10.20)	Green	549
4	[Mn(C <sub>18</sub> H <sub>16</sub> N <sub>4</sub> O <sub>2</sub> )(OAc)](OAc) <sub>2</sub>	9.89 (9.96)	52.26 (52.17)	4.49 (4.52)	10.27 (10.14)	Off white	552
5	[Fe(C <sub>18</sub> H <sub>16</sub> N <sub>4</sub> O <sub>2</sub> )Cl]Cl <sub>2</sub>	11.64 (11.55)	44.83 (44.76)	3.29 (3.31)	11.54 (11.60)	Dark green	482.5
6	[Fe(C <sub>18</sub> H <sub>16</sub> N <sub>4</sub> O <sub>2</sub> )(NO <sub>3</sub> )](NO <sub>3</sub> ) <sub>2</sub>	9.87 (9.96)	38.39 (38.43)	2.93 (2.84)	17.21 (17.43)	Dark red	562
7	[Fe(C <sub>18</sub> H <sub>16</sub> N <sub>4</sub> O <sub>2</sub> )(OAc)](OAc) <sub>2</sub>	10.19 (10.12)	52.12 (52.07)	4.64 (4.52)	10.18 (10.12)	Yellow	553

concentrated HNO<sub>3</sub>, indicating their presence inside as well as outside the coordination sphere. Conductivity measurements in DMSO indicated them to be electrolytic in nature (140–150 ohm<sup>-1</sup> cm<sup>2</sup> mol<sup>-1</sup>)<sup>15</sup>. All compounds gave satisfactory elemental analysis results, as shown in Table 1.

### IR spectra

In the infrared spectrum of succinyldihydrazide, a pair of bands corresponding to  $\nu(\text{NH}_2)$  is present at  $\sim 3200$  cm<sup>-1</sup> and  $\sim 3250$  cm<sup>-1</sup>, but this was absent in the IR spectra of all the complexes<sup>16</sup>. However, a single, broad, medium band at  $\sim 3350$ – $3400$  cm<sup>-1</sup> was observed in the spectra of all the complexes, which may be assigned to  $\nu(\text{NH})$ <sup>17,18</sup>. A strong peak at  $\sim 1665$  cm<sup>-1</sup> in the IR spectrum of succinyldihydrazide is assigned to the  $>\text{C}=\text{O}$  group of the CONH moiety. This peak was shifted to a lower frequency ( $\sim 1625$ – $1640$  cm<sup>-1</sup>) in the spectra of all the complexes<sup>19,20</sup>, suggesting coordination of the oxygen of the carbonyl group with the metal. Further, no strong absorption band was observed near  $1700$  cm<sup>-1</sup> in the IR spectra of all complexes, as observed in the spectrum of benzil. This indicates the absence of  $>\text{C}=\text{O}$  groups of the benzil moiety in the complexes. These facts confirm the condensation of carbonyl groups of the benzil and amino groups of succinyldihydrazide<sup>21,22</sup>. The IR spectra of the complexes showed a new strong absorption band in the region  $\sim 1590$ – $1610$  cm<sup>-1</sup>, which may be attributed to  $\nu(\text{C}=\text{N})$ <sup>23,24</sup>. These results provide strong evidence for the formation of a macrocyclic frame<sup>25</sup>. The lower value of  $\nu(\text{C}=\text{N})$  indicates coordination of the nitrogens of azomethine and the metal<sup>26</sup>. The bands present at  $\sim 1350$ – $1000$  cm<sup>-1</sup> are assigned to  $\nu(\text{C}-\text{N})$  vibration. The bands present at  $\sim 3040$  cm<sup>-1</sup> may be assigned to  $\nu(\text{C}-\text{H})$  vibrations of the benzil moiety.

### Far infrared spectra

The far infrared spectra showed bands in the region  $\sim 425$ – $445$  cm<sup>-1</sup> corresponding to  $\nu(\text{M}-\text{N})$  vibrations in all the complexes<sup>27–29</sup>. The presence of a band in all the complexes at the  $\sim 425$ – $445$  cm<sup>-1</sup> region originates from (M–N) azomethine vibration modes and gives an idea about coordination of azomethine nitrogens with the metal<sup>30</sup>. The bands present at  $\sim 300$ – $315$  cm<sup>-1</sup> are due to  $\nu(\text{M}-\text{Cl})$ <sup>27,29</sup> and bands present at  $\sim 220$ – $250$  cm<sup>-1</sup> in all nitrate complexes to  $\nu(\text{M}-\text{O})$ <sup>27</sup>.

### Magnetic measurements and electronic spectra

**Chromium complexes** Magnetic moments of chromium complexes were found in the range of 4.25–4.60 BM. The

electronic spectra of chromium complexes showed bands at  $\sim 9010$ – $9260$ ,  $13,000$ – $13,550$ ,  $17,460$ – $18,300$ ,  $27,400$ – $27,800$ , and  $34,850$  cm<sup>-1</sup>. However, these spectral bands cannot be interpreted in terms of four or six coordinated environments around the metal atom. In turn, the spectra are comparable to those of five-coordinated Cr(III) complexes, whose structures have been confirmed with the help of X-ray measurements<sup>31</sup>. Thus, keeping in view the analytical data and electrolytic nature of these complexes, a five-coordinated square pyramidal geometry may be assigned to these complexes. Thus, assuming the symmetry C<sub>4v</sub> for these complexes<sup>32</sup>, the various spectral bands may be assigned as:  ${}^4\text{B}_1 \rightarrow {}^4\text{E}^a$ ,  ${}^4\text{B}_1 \rightarrow {}^4\text{B}_2$ ,  ${}^4\text{B}_1 \rightarrow {}^4\text{A}_2$ , and  ${}^4\text{B}_1 \rightarrow {}^4\text{E}^b$ .

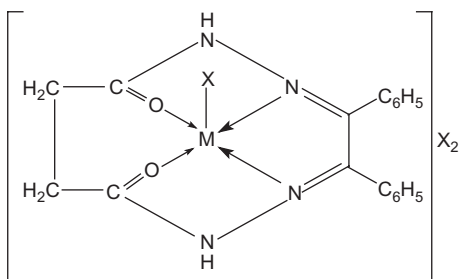
**Manganese complex** The magnetic moment of the manganese complex was found to be 4.80 BM. The electronic spectrum of the manganese complex showed three d–d bands which lay in the regions  $12,200$ ,  $16,100$ , and  $35,400$  cm<sup>-1</sup>. The higher energy band at  $35,400$  cm<sup>-1</sup> may be assigned to charge transfer transitions. The spectrum resembled those reported for five-coordinate square pyramidal manganese porphyrins<sup>32</sup>. This idea is further supported by the presence of the broad-ligand field band at  $20,400$  cm<sup>-1</sup>, diagnostic of C<sub>4v</sub> symmetry, and thus the various bands may be assigned as follows:  ${}^5\text{B}_1 \rightarrow {}^5\text{A}_1$ ,  ${}^5\text{B}_1 \rightarrow {}^5\text{B}_2$ , and  ${}^5\text{B}_1 \rightarrow {}^5\text{E}$ , respectively. The band assignment in single electron transition may be made as:  $d_{z^2} \rightarrow d_{x^2-y^2}$ ,  $d_{xy} \rightarrow d_{x^2-y^2}$ , and  $d_{xy}, d_{yz} \rightarrow d_{x^2-y^2}$ , respectively, in order of increasing energy. However, the complexes do not have idealized C<sub>4v</sub> symmetry.

**Iron complexes** The magnetic moments of the iron complexes lay in the range 5.80–5.92 BM. The electronic spectra of the trivalent iron complexes showed various bands,  $9800$ – $9900$ ,  $15,500$ – $15,550$ ,  $27,600$ – $27,700$  cm<sup>-1</sup>, and these bands do not suggest octahedral or tetrahedral geometry around the metal atom. The spectral bands are consistent with the range of those reported for five-coordinate square pyramidal iron(III) complexes<sup>33</sup>. Assuming C<sub>4v</sub> symmetry for these complexes, the various bands can be assigned as:  $d_{xy} \rightarrow d_{xz}$ ,  $d_{yz}$  and  $d_{xy} \rightarrow d_{z^2}$ . Any attempt to make an accurate assignment is difficult due to interactions of the metal–ligand pi-bond systems lifting the degeneracy of the d<sub>xz</sub> and d<sub>yz</sub> pair.

Based on elemental analyses, conductivity and magnetic measurements, and electronic, IR and far IR spectral studies, the general structure shown in Figure 2 may be proposed for these complexes.

### Biology

The synthesized macrocyclic complexes were tested for their *in vitro* antibacterial activity against four test bacteria, *Bacillus cereus* (MTCC 1272), *Salmonella typhi* (MTCC 733), *Escherichia coli* (MTCC 739), and *Staphylococcus aureus* (MTCC 1144). The MIC (minimum inhibitory concentration) shown by the complexes against these bacterial strains was compared with the MIC shown by standard antibiotics linezolid and cefaclor (Table 2 and Figure 3). Complex 1 showed



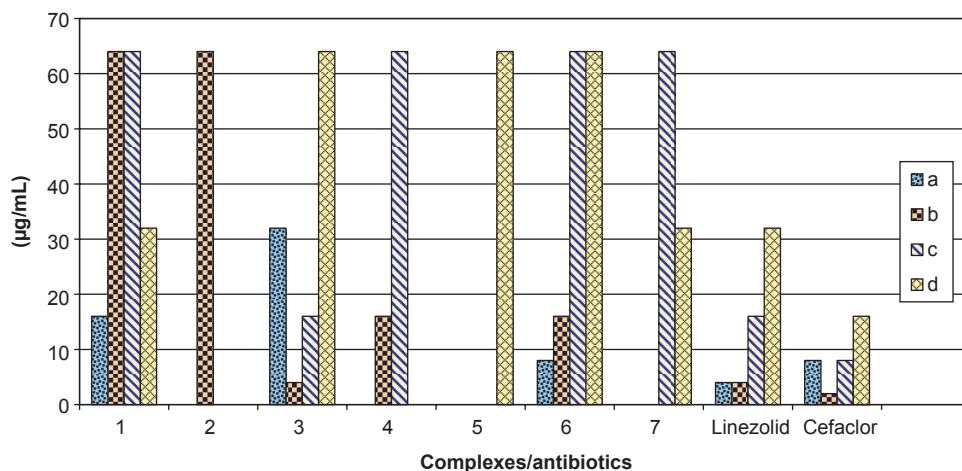
**Figure 2.** General structure of the complexes, where M = Cr(III), Mn(III), Fe(III), X = Cl<sup>-</sup>, NO<sub>3</sub><sup>-</sup>, CH<sub>3</sub>COO<sup>-</sup> for Cr(III) and Fe(III) and X = CH<sub>3</sub>COO<sup>-</sup> for Mn(III).

a minimum inhibitory concentration of 32 µg/mL against bacterial strain *Salmonella typhi* (MTCC 733), which is equal to the MIC shown by standard antibiotic linezolid against the same bacterial strain. Complex 3 registered a minimum inhibitory concentration of 4 µg/mL against bacterial strain *Staphylococcus aureus* (MTCC 1144), which is equal to the MIC shown by standard antibiotic linezolid against the same bacterial strain. Further, it showed a minimum inhibitory concentration of 16 µg/mL against bacterial strain *Escherichia coli* (MTCC 739), which is equal to the MIC shown by standard antibiotic linezolid against the same bacterial strain. Complex 4 registered a minimum inhibitory concentration ranging from 16 to 64 µg/mL. Complex 6 registered a minimum inhibitory concentration of 8 µg/mL against the bacterial strain *Bacillus cereus* (MTCC 1272), which is equal to the MIC shown by standard antibiotic cefaclor against the same bacterial strain. Complex 7 registered a minimum inhibitory concentration of 32 µg/mL against bacterial strain *Salmonella typhi* (MTCC 733), which is equal to the MIC shown by standard antibiotic linezolid against the same bacterial strain. Among the series under test for determination of the minimum inhibitory concentration, complex 3 was found to be the most potent.

**Table 2.** Minimum inhibitory concentration (MIC) shown by complexes against test bacteria using agar dilution assay.

Series no.	Complex	MIC (µg/mL)			
		a	b	c	d
1	[Cr(C <sub>18</sub> H <sub>16</sub> N <sub>4</sub> O <sub>2</sub> )Cl]Cl <sub>2</sub>	16	64	64	32
2	[Cr(C <sub>18</sub> H <sub>16</sub> N <sub>4</sub> O <sub>2</sub> )(NO <sub>3</sub> ) <sub>2</sub> ](NO <sub>3</sub> ) <sub>2</sub>	128	64	128	128
3	[Cr(C <sub>18</sub> H <sub>16</sub> N <sub>4</sub> O <sub>2</sub> )(OAc)](OAc) <sub>2</sub>	32	4	16	64
4	[Mn(C <sub>18</sub> H <sub>16</sub> N <sub>4</sub> O <sub>2</sub> )(OAc)](OAc) <sub>2</sub>	—	16	64	>128
5	[Fe(C <sub>18</sub> H <sub>16</sub> N <sub>4</sub> O <sub>2</sub> )Cl]Cl <sub>2</sub>	—	128	>128	64
6	[Fe(C <sub>18</sub> H <sub>16</sub> N <sub>4</sub> O <sub>2</sub> )(NO <sub>3</sub> ) <sub>2</sub> ](NO <sub>3</sub> ) <sub>2</sub>	8	16	64	64
7	[Fe(C <sub>18</sub> H <sub>16</sub> N <sub>4</sub> O <sub>2</sub> )(OAc)](OAc) <sub>2</sub>	128	—	64	32
	Cefaclor	8	2	8	16
	Linezolid	4	4	16	32

Note. —, no activity; a, *Bacillus cereus* (MTCC 1272); b, *Staphylococcus aureus* (MTCC 1144); c, *Escherichia coli* (MTCC 739); d, *Salmonella typhi* (MTCC 733); cefaclor and linezolid are standard antibiotics.



**Figure 3.** Comparison of minimum inhibitory concentration (MIC) of complexes with those of standard antibiotics up to concentration of 64 µg/mL. a, *Bacillus cereus* (MTCC 1272); b, *Staphylococcus aureus* (MTCC 1144); c, *Escherichia coli* (MTCC 739); d, *Salmonella typhi* (MTCC 733); cefaclor and linezolid are standard antibiotics.

However, complexes 2, 4, and 5 showed poor antibacterial activity or no activity against all bacterial strains among the whole series.

## Conclusions

Based on elemental analyses, conductivity and magnetic measurements, and electronic, IR, and far IR spectral studies, the structure as shown in Figure 2 may be proposed for these complexes.

It has been suggested that chelation/coordination reduces the polarity of the metal ion mainly because of partial sharing of its positive charge with the donor group within the whole chelate ring system. This process of chelation thus increases the lipophilic nature of the central metal atom, which in turn favors its permeation through the lipid layer of the membrane, thus causing the metal complex to cross the bacterial membrane more effectively and hence increase the activity of the complex. Aside from this, many other factors such as solubility, dipole moment, and conductivity influenced by the metal ion may be possible reasons for the remarkable antibacterial activities of these complexes<sup>34-37</sup>. It also has been observed that some moieties, such as the azomethine linkage or heteroaromatic nucleus, introduced into such compounds exhibit extensive biological activities that may be responsible for the increase in hydrophobic character and liposolubility of the molecules in crossing the cell membrane of the microorganism, thus enhancing the biological utilization ratio and activity of complexes<sup>38,39</sup>.

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**Declaration of interest:** The authors report no conflicts of interest.

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